



Short Communication

PANAXYNOL, A POLYACETYLENE COMPOUND ISOLATED FROM ORIENTAL MEDICINES, INHIBITS MAMMALIAN LIPOXYGENASES

JUHA ALANKO,* YUKO KURAHASHI, TANIHIRO YOSHIMOTO,
 SHOZO YAMAMOTO† and KIMIYE BABA‡

Department of Biochemistry, Tokushima University, School of Medicine, Kuramoto-cho, Tokushima 770; ‡Osaka University of Pharmaceutical Sciences, Kawai 2-10-65, Matsubara, Osaka 580, Japan

(Received 14 January 1994; accepted 8 August 1994)

Abstract—Panaxynol is a polyacetylene compound isolated from commonly used oriental medicines, and its effects on various cyclooxygenases and lipoxygenases were investigated. The compound had only a marginal effect on cyclooxygenase activities (IC_{50} values $\geq 100 \mu M$), but inhibited lipoxygenases; 5-lipoxygenase (IC_{50} , $2 \mu M$), two isoforms of 12-lipoxygenase (leukocyte-type, $1 \mu M$; platelet-type, $67 \mu M$) and 15-lipoxygenase ($4 \mu M$). Thus, panaxynol inhibited leukocyte-type 12-lipoxygenase much more effectively than platelet-type 12-lipoxygenase. Fincarindiol, an analogue of panaxynol, inhibited these lipoxygenases with higher IC_{50} values than panaxynol. These compounds could provide a clue to develop a selective inhibitor of one isoform of 12-lipoxygenase.

Key words: arachidonic acid; cyclooxygenase; fincarindiol; platelets; leukocytes

Panaxynol [(9Z)-heptadeca-1,9-dien-4,6-diyn-3-ol] was isolated from Ginseng radix, “Fang-Feng” and Panax ginseng [1, 2] and fincarindiol [(9Z)-heptadeca-1,9-dien-4,6-diyn-3,8-diol] from “Fang-Feng” [1] (Fig. 1). They are commonly used oriental medicines. Anti-inflammatory and anti-platelet-aggregatory actions of panaxynol were previously reported [1, 2]. In view of their 1,9-dien-4,6-diyn structure as an analogue of the polyen structure of arachidonic acid, we investigated their effects on various mammalian cyclooxygenases and lipoxygenases. Cyclooxygenase incorporates two molecules of oxygen into arachidonic acid, and initiates the biosynthesis of prostaglandins and thromboxanes [3]. Lipoxygenase incorporates only one molecule of oxygen at particular positions of unsaturated fatty acid [4, 5]. Biologically potent leukotrienes are synthesized via the 5-lipoxygenase pathway of arachidonic acid [6]. 15-Lipoxygenase was found in leukocytes [7] and reticulocytes [8]. There are two types of 12-lipoxygenase (platelet-type and leukocyte-type), which are distinguishable in terms of substrate specificity, immunogenicity and amino acid sequence [4, 5].

Materials and Methods

[1- ^{14}C]Arachidonic acid (2.04 GBq/mmol) was purchased from Amersham International (Amersham, U.K.), and unlabeled arachidonic acid from Nu-Chek Prep (Elysian, MN, U.S.A.). Panaxynol and fincarindiol were isolated and purified from Saposhnikovia radix as described previously [1]. Sheep cyclooxygenase purified from seminal vesicle by immunoaffinity chromatography was supplied by K. Yamamoto of this laboratory. A suspension of platelet microsomes was used as human cyclooxygenase. Porcine 5-lipoxygenase [9] and 12-lipoxygenase [10] were purified from leukocytes by immunoaffinity chromatography as described. Recombinant 12-lipoxygenase of human platelets

was prepared by M. Nakamura and K. Kishimoto of this laboratory. 15-Lipoxygenase was a lysate from rabbit reticulocytes.

The standard assay mixtures (200 μL) were as follows: for cyclooxygenase 0.1 M Tris-HCl buffer (pH 8.0), 2 μM hematin and 5 mM tryptophan; for 12- and 15-lipoxygenases 0.05 M Tris-HCl buffer (pH 7.5); and for 5-lipoxygenase 0.05 M potassium phosphate buffer (pH 7.4), 2 mM $CaCl_2$ and 2 mM ATP. The enzyme amount was adjusted so that approx. 50% of arachidonic acid could be oxygenated. The reaction was started by the addition of [1- ^{14}C]arachidonic acid (100,000 cpm/5 nmol in 5 μL of ethanol). Cyclooxygenase reaction was performed at 24° for 1 min, and lipoxygenase reactions at 30° for 5 min. Incubation was stopped by adding 300 μL ice-cold diethyl ether/methanol/0.2 M citrate (30:4:1, by vol.). Thin layer chromatography was performed as described previously with a solvent system of diethyl ether/petroleum ether/acetic acid (85:15:0.1 by vol.) [9]. Panaxynol and fincarindiol were added in 4 μL of dimethyl sulfoxide.

Results and Discussion

Panaxynol had essentially no effect on ovine and human cyclooxygenases up to 30 μM concentration except for a slight activation of the enzyme activity at lower concentrations (Fig. 2). IC_{50} values were higher than 100 μM . In contrast, panaxynol inhibited various lipoxygenases (Fig. 2). IC_{50} values for 5-lipoxygenase, 12-lipoxygenases of leukocyte-type and platelet-type and 15-lipoxygenase were 2, 1, 67 and 4 μM , respectively. The most potent inhibition was observed with leukocyte-type 12-lipoxygenase, and the compound was slightly less effective on 5- and 15-lipoxygenases. It should be noted that platelet-type 12-lipoxygenase required approx. 70 times higher concentration of the inhibitor than leukocyte-type 12-lipoxygenase.

As described above, cyclooxygenases and lipoxygenases were distinguished in terms of the inhibition by panaxynol. However, panaxynol is not a selective inhibitor for a particular lipoxygenase, inhibiting as it does 5-, 12- and 15-lipoxygenases. Two 12-lipoxygenases of porcine leukocytes and human platelets were inhibited by panaxynol with IC_{50}

† Corresponding author. Tel. 81-886-31-3111, ext. 2220; FAX 81-886-33-6409.

* On leave from the Department of Biomedical Sciences, University of Tampere, Finland.

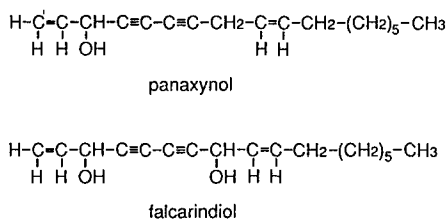


Fig. 1. Structures of panaxynol and falcarindiol.

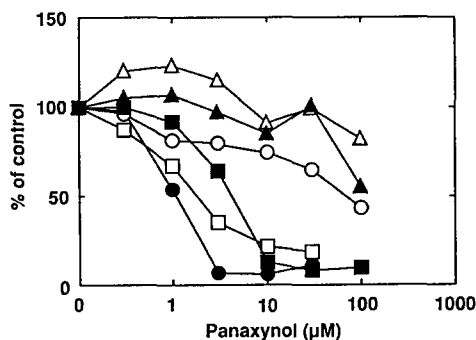


Fig. 2. Effects of panaxynol on various preparations of cyclooxygenase and lipoxygenase. The following amounts of enzymes were assayed with arachidonic acid under the standard conditions; cyclooxygenase of ovine seminal vesicle (open triangles, 1.0 μg protein), cyclooxygenase of human platelets (closed triangles, 148 μg protein), 5-lipoxygenase of porcine leukocytes (open squares, 8.0 μg protein), 12-lipoxygenase of porcine leukocytes (closed circles, 2.0 μg protein), recombinant 12-lipoxygenase of human platelets (open circles, 0.7 μg protein) and 15-lipoxygenase of rabbit reticulocytes (closed squares, 16.1 μg protein). Mean of four to five experiments.

values different by almost two orders of magnitude. This finding would provide a clue to develop a selective inhibitor to distinguish the two types of 12-lipoxygenase. Such a selective 12-lipoxygenase inhibitor would help to elucidate still unknown physiological roles of 12-lipoxygenases.

Falcarindiol is an analogue of panaxynol with an additional hydroxyl group at 8 position (Fig. 1). The compound was a less effective inhibitor of the lipoxygenases tested above. IC_{50} values for 5-lipoxygenase, 12-lipoxygenase of leukocyte-type and platelet-type and 15-lipoxygenase were 7, 48, >100 and 18 μM , respectively. Falcarindiol had essentially no effect on either ovine or human cyclooxygenases up to 30 μM concentration.

Eicosa-5,8,11,14-tetraenoic acid was earlier shown to inactivate cyclooxygenase [11] and lipoxygenase [11, 12] by its covalent binding to the enzyme proteins. Given its 1,9-dien-4,6-diyn structure, panaxynol was subjected to a kinetic study with porcine leukocyte 12-lipoxygenase which was inhibited most potently by the compound. Since the enzyme is known as a suicide enzyme and the reaction time course is almost linear for only 1 min or so, the enzyme reaction was performed for 1 min. Panaxynol was shown to be a competitive inhibitor as examined by Lineweaver-Burk plots. Such a competitive nature suggests that panaxynol is bound to the active site of the enzyme as a

substrate analogue. However, it is possible that the inhibitor is covalently bound to the enzyme protein by its diyn portion. Preincubation of porcine leukocyte 12-lipoxygenase with panaxynol was carried out on ice because of the enzyme lability. IC_{50} values after 30 and 60 min preincubations were 0.7 and 0.5 μM , respectively, as compared with 1 μM after 1 min preincubation. The enzyme activity decreased by 60 and 76% after incubation for 30 and 60 min. It is not certain whether such a slow inactivation may be attributed to a slow covalent binding of the inhibitor to the enzyme protein. First, panaxynol interacts with the active site of 12-lipoxygenase competing with arachidonic acid, and then may be gradually covalently bound to the enzyme protein to be an irreversible inhibitor.

As examined previously with whole platelet cells, the production of thromboxane B_2 [1, 2] and 12-hydroxy-5,8,10-heptadecatrienoic acid [2] was inhibited by panaxynol at 0.4–0.8 mM [1] and 0.08–0.8 mM [2]. At such high concentrations cyclooxygenase activity may be inhibited by panaxynol, resulting in the reduced production of thromboxane B_2 . The lipoxygenase inhibition observed in the present work required much lower concentrations of panaxynol.

Acknowledgements—Juha Alanko was a visiting scientist at Tokushima University under the agreement between the Japan Society for Promotion of Science and the Academy of Finland. This work was supported by grants-in-aid from the Ministry of Education, Science and Culture of Japan, the Japanese Foundation of Metabolism and Diseases, the Ono Medical Research Foundation, the CIBA-GEIGY Foundation for the Promotion of Science, and the Japan Foundation for Applied Enzymology.

REFERENCES

- Baba K, Tabata Y, Kozawa M, Kimura Y and Arichi S, Studies on Chinese traditional medicine "Fang-Feng" (I) Structures and physiological activities of polyacetylene compounds from *Saposhnikovia radix*. *Shoyakugaku Zasshi* 41: 189–194, 1987.
- Teng C-M, Kuo S-C, Ko F-N, Lee J-C, Lee L-G, Chen S-C and Huang T-F, Antiplatelet actions of panaxynol and ginsenosides isolated from ginseng. *Biochim Biophys Acta* 990: 315–320, 1989.
- Yamamoto S, Enzymes in the arachidonic acid cascade. In: *Prostaglandins and Related Substances* Eds. (Pace-Asciak C and Granström E), 171–202. Elsevier Science Publishers, Amsterdam, 1983.
- Yamamoto S, "Enzymatic" lipid peroxidation: reactions of mammalian lipoxygenases. *Free Rad Biol Med* 10: 149–159, 1991.
- Yamamoto S, Mammalian lipoxygenases: molecular structures and functions. *Biochim Biophys Acta* 1128: 117–131, 1992.
- Samuelsson B, Leukotrienes: mediators of immediate hypersensitivity reactions and inflammation. *Science* 220: 568–575, 1983.
- Narumiya S, Salmon JA, Cottee FH, Weatherley BC and Flower RJ, Arachidonic acid 15-lipoxygenase from rabbit peritoneal polymorphonuclear leukocytes. Partial purification and properties. *J Biol Chem* 256: 9583–9592, 1981.
- Rapoport SM, Schewe T, Wiesner R, Halangk W, Ludwig P, Janicke-Höhn M, Tannert C, Hiebsch C and Klatt D, The lipoxygenase of reticulocytes. Purification, characterization and biological dynamics of the lipoxygenase; its identity with the respiratory inhibitors of the reticulocyte. *Eur J Biochem* 96: 545–561, 1979.
- Ueda N, Kaneko S, Yoshimoto T and Yamamoto S, Purification of arachidonate 5-lipoxygenase from porcine leukocytes and its reactivity with hydro-

- peroxyeicosatetraenoic acid. *J Biol Chem* **261**: 7982–7988, 1986.
10. Yokoyama C, Shinjo F, Yoshimoto T, Yamamoto S, Oates JA and Brash AR, Arachidonate 12-lipoxygenase purified from porcine leukocytes by immunoaffinity chromatography and its reactivity with hydroperoxyeicosatetraenoic acid. *J Biol Chem* **261**: 16714–16721, 1986.
 11. Downing DT, Ahern DG and Bachta M, Enzyme inhibition by acetylenic compounds. *Biochem Biophys Res Commun* **40**: 218–223, 1970.
 12. Bokoch GM and Reed PW, Evidence for inhibition of leukotriene A₄ synthesis by 5,8,11,14-eicosatetraenoic acid in guinea pig polymorphonuclear leukocytes. *J Biol Chem* **256**: 4156–4159, 1981.